View Article Online / Journal Homepage / Table of Contents for this issue
Dynamic Article Links

Catalysis
Science & Technology

Cite this: *Catal. Sci. Technol.*, 2011, **1**, 1423–1431

www.rsc.org/catalysis

PAPER

The activity of invertase immobilized on cashew nut shell liquid-templated large pore silica hybrids

Egid B. Mubofu,* James E. G. Mdoe and Grace Kinunda

Received 27th January 2011, Accepted 6th July 2011
DOI: 10.1039/c1cy00033k

Published on 22 August 2011 on http://pubs.rsc.org | doi:10.1039/C1CY00033K
Downloaded on 13 February 2013

The characterization and activity of invertase enzymes immobilized on large pore micelle templated silica (LP-MTS) hybrid materials is reported. The LP-MTS hybrid materials were prepared by a co-condensation of tetraethoxysilane (TEOS) and 3-aminopropyltrimethoxysilane (AMPTS) in a cashew nut shell liquid (CNSL) template. A commercially available dodecylamine template was also used to afford similar materials, hereinafter abbreviated as DDA-MTS, for comparison purposes. The prepared materials were characterized by different techniques to determine their physicochemical properties. The maximum loading for the amine groups in LP-MTS and DDA-MTS were 3.3 and 2.8 mmol g^{-1}, respectively. Modification of the materials for immobilization of the invertase enzyme was done by reacting them with glutaraldehyde resulting in Glu-LP-MTS or Glu-DDA-MTS. The as-prepared hybrid materials have surface areas ranging from 100 to 214 m^2 g^{-1} with pore diameters ranging from 3.1 to 25 nm. Scanning electron microscopy (SEM) images show that LP-MTS and DDA-MTS materials comprise of roughly spherical particles whereas enzyme and glutaraldehyde supported micelle templated silicas show a rupture of the spherical particles to a fine powder. The activities of free and immobilized invertases have been determined by measuring the amount of reducing sugar produced upon hydrolysis of sucrose at different temperatures, pH and substrate concentrations. Both free and immobilized invertase enzymes showed a maximum activity at a particular optimum temperature, pH and substrate concentration. The maximum activity for the free invertase was 15 229 U at pH 5.0 and at 40 °C whereas those of the LP-MTS immobilized invertases were 14 833 U and 14 625 U for covalent and cross-linked invertases, respectively, at pH 4.0 and 40 °C. The maximum activity for DDA-MTS covalently immobilized invertase was 4750 U, at pH 4.0 and 40 °C.

Introduction

The application of enzymes as biocatalysts is important in industrial applications, biosensors and drugs.[1–6] Unlike metal catalysts, enzymes are highly desirable catalysts when the specificity of the reaction, mild reaction conditions and environmental restrictions are major issues. Despite advantages, the use of enzymes on an industrial scale is hampered by several factors, including the high cost of enzymes and their instability. Furthermore, extreme reaction conditions inactivate enzymes through denaturation due to either changes of conformation or other transformations in their chemical structure. In addition, the utilization of natural enzymes has other processing difficulties such as the problem on reuse of enzymes and product contamination.[7–9] Enzymes are soluble in aqueous media and it is therefore difficult and expensive to recover them at the end of the catalytic process. As a result, native enzymes often suffer severe limitations in broader industrial applications. One of the approaches to resolve these difficulties is to immobilize them on solid surfaces which can produce recoverable and stable heterogeneous biocatalysts. Enzyme immobilization with different kinds of matrices have been investigated by several workers.[9–13] The fixation of an enzyme into inorganic materials combines the selectivity of the enzyme with improved chemical and mechanical stability of the enzyme. The storage properties of enzymes are also improved by immobilization.

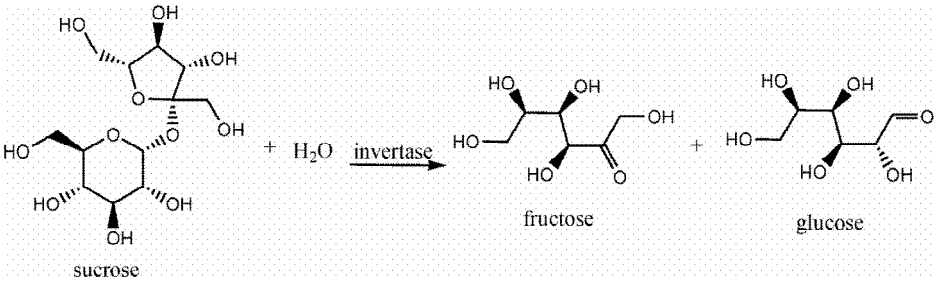
Mesoporous silicas have been the subject of intense study[5–29] because they offer attractive supports for a range of catalytic complexes due to their wide range of topologies and connectivities with dimensions of up to 3 nm and beyond. However, due to small pore size and non-open pore hindrance, the immobilized enzymes have sometimes shown lower activity than free enzymes, and the non-uniform pore sizes of most silica gel supports make the processes less reproducible. Micelle templated silica (MTS) materials have tunable and

Department of Chemistry, University of Dar es Salaam, P.O. Box 35061, Tanzania. E-mail: ebmubofu@gmail.com; Fax: +255-22-2410038; Tel: +255-22-2410038

Published on 22 August 2011 on http://pubs.rsc.org | doi:10.1039/C1CY00033K
Downloaded on 13 February 2013

functionalizable surfaces with restricted nanospaces that are ideal for enzyme immobilization. Recently, many research groups have immobilized enzymes on MTS which showed improvements on stability, activity, product specificity, and resistance to extreme environmental conditions.[6,18,19,26–28] MTS supports show biocompatibility, have a low cytotoxicity, large surface areas, and can be easily functionalized and hence are also attractive for biomedical applications.[20–25]

Invertase is a widely used enzyme in the food industry particularly in the production of sweeteners used in beverages, jams, and artificial honey[30] as it catalyzes the cleaving of the α-1,4 glycosidic bonds of sucrose to produce glucose and fructose.



The enzyme exhibits relatively high activity over a broad range of pH (3.5–5.5) with the optimum[31] near pH 4.5 and its activity reaches maximum at about 55 °C. Due to its potential in industries, availability and ease of handling, invertase has been immobilized on a number of carriers.[32–38] However, most of these supports are based on soft gels which have low mechanical strength and in some cases low thermal and chemical stability.[34] Organic–inorganic mesoporous silica materials are most promising because they offer tunability and most of the required properties of a good enzyme support. A good number of the mesoporous materials that have been studied for bioimmobilization posses a relatively small pore diameter not exceeding 10 nm.[1–10,26–28] This pore diameter is smaller than most biomolecules, which are more than 10 kD. Large pore micelle templated silicas (LP-MTS) with pore dimensions in the range of 17 nm to 25 nm are potentially attractive supports for bioimmobilization due to their large pore dimensions. LP-MTS with pore dimensions in this range have previously been prepared in our group using a cashew nut shell liquid (CNSL) template.[29] Herein, we report the immobilization protocol, the systemic characterization and catalytic testing of LP-MTS immobilized invertase. For comparison, invertase immobilized on MTS prepared from the commercially available surfactant, dodecylamine (DDA) is also reported.

Results and discussion

Porosity properties of LP-MTS/DDA-MTS and supported glutaraldehyde

The porosity properties of the LP-MTS/DDA-MTS and their derivatives were determined by analysis of nitrogen adsorption isotherms using the BET method. Table 1 gives a summary of the BET surface areas, pore volumes and average pore diameters of these materials.

The LP-MTS and modified LP-MTS materials display surface areas ranging from 100 to 190 $m^2 g^{-1}$ with pore diameters ranging from 11 to 25 nm. On the other hand, the DDA-MTS and Glu-DDA-MTS have surface areas of 214 and 212 $m^2 g^{-1}$, respectively, both having pore diameters of 3.1 nm. The pore diameters and pore volumes for LP-MTS decreased upon modification whereas in DDA-MTS it remained the same. This shows that for DDA-MTS, the modification did not take place inside the pores, rather on the void spaces of the particles. The slight decrease in surface areas and pore diameters observed after each step of MTS modification is due to steric effects caused by the functional groups introduced into the interior of the pores which have reduced the area available for nitrogen physisorption and this implies that the modification for LP-MTS takes place inside the pores.[39,40]

The shapes of the isotherms obtained for all materials are type IV (Fig. 1). This is a typical shape for mesoporous materials with initial gradual increase of nitrogen uptake at low relative pressure (<0.1). Above that, the isotherm forms a plateau followed by another sharp rise of nitrogen uptake starting from about P/P_o = 0.8.

The initial gradual increase of nitrogen uptake at low relative pressure (<0.1) observed in all isotherms is attributed to the formation of nitrogen monolayer on the adsorbent surface. There is a sharp rise of nitrogen uptake from P/P_o = 0.8 for these materials and this behavior can be associated with the formation of multilayer of the adsorbate, causing capillary condensation in the pores.[40–44] A slightly low nitrogen uptake at the same relative pressure is observed in the LP-MTS derivatives. This implies that materials introduced during modification of LP-MTS to form Glu-LP-MTS and the immobilized invertase occupied some parts of the

Table 1 Physical-chemical properties of LP-MTS/DDA-MTS, Glu-LP-MTS/Glu-DDA-MTS, covalent immobilized invertase, cross linked invertase and free invertase

Material	BET Surface area ($m^2 g^{-1}$)	Average pore diameter (nm)	Total pore volume ($cm^3 g^{-1}$)
LP-MTS	160	24.8	2.9
DDA-MTS	214	3.1	0.1
Glu-LP-MTS	189	11.7	2.4
Glu-DDA-MTS	212	3.1	0.1
Covalent immobilized invertase on LP-MTS	116	11.5	0.6
Cross-linked invertase on LP-MTS	123	11.5	0.7
Free invertase	30	3.1	0.4

This journal is © The Royal Society of Chemistry 2011

Published on 22 August 2011 on http://pubs.rsc.org | doi:10.1039/C1CY00033K
Downloaded on 13 February 2013

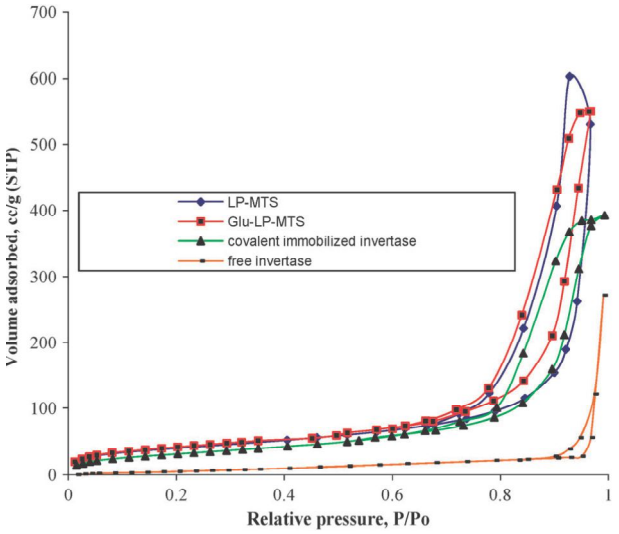


Fig. 1 Nitrogen adsorption–desorption isotherms for LP-MTS, Glu-LP-MTS, covalent immobilized invertase and free invertase.

LP-MTS pore volume. The isotherms of LP-MTS and that of Glu-LP-MTS are similar in that they rise rapidly towards $P/P_o = 1$. On the other hand, the isotherms for the immobilized invertases are similar. Their curves tend to be horizontal towards $P/P_o = 1$ and show mesoporosity characteristics. The significant difference between the isotherms of LP-MTS, Glu-LP-MTS and that of immobilized invertase lies in the amount of nitrogen adsorbed at the same relative pressure. The Glu-LP-MTS isotherm and that of immobilized invertase show slightly low nitrogen uptake compared to the LP-MTS support. The isotherm of LP-MTS and its derivatives clearly show hysteresis loops at a relative pressure above 0.6 whereas free invertase shows a hysteresis loop at a relative pressure above 0.8. The volume of nitrogen adsorbed depends on the pore diameter, pore volume and surface area of the material. A large amount of nitrogen is adsorbed on the parent LP-MTS material and decreases as its modification takes place. This is in line with the decrease in pore sizes and surface areas upon functionalization of the support.

The adsorption–desorption isotherms of MTS prepared by DDA templates are represented in Fig. 2.

The isotherms have two substantial initial inflexion points due to sharp nitrogen uptakes; the first is at low relative

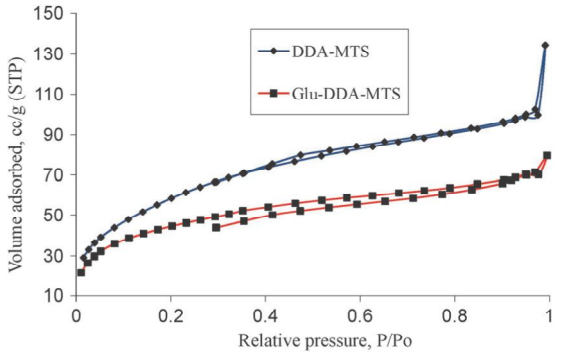


Fig. 2 Nitrogen adsorption–desorption isotherms for DDA-MTS and Glu-DDA-MTS.

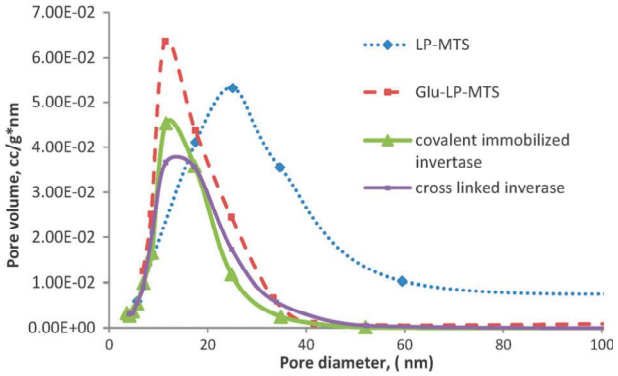


Fig. 3 Pore size distributions of LP-MTS, Glu-LP-MTS, covalent immobilized invertase and cross-linked invertase.

pressure (<0.1) and the second is at a relative pressure of about 0.9 with some hysteresis loops at a relative pressure of about 0.4. The higher nitrogen uptake at low relative pressure is indicative of monolayer adsorption of these materials. The Glu-DDA-MTS adsorbs less nitrogen relative to DDA-MTS at a given relative pressure.

The pore size distributions of LP-MTS, Glu-LP-MTS and immobilized invertases show a wide pore size distribution with the majority of pores ranging from 10 nm to 40 nm as observed in Fig. 3. There is a decrease in the pore size distributions upon fuctionalization of LP-MTS to Glu-LP-MTS. In contrast, the pore size distribution of DDA based materials is narrow with the average pore diameters of 3 nm (Table 1).

The pore size distributions for the LP-MTS are generally wide. The majority of the pores have diameters ranging from 11 to 25 nm and an average pore diameter of 18 nm. Unlike LP-MTS supports, DDA-MTS supports have a narrow pore size distribution. The reason for the unexpectedly large pore diameters observed in CNSL-based hybrids is still unclear, and further studies on the templating mechanism are underway. It was noted that, the greater the modification of LP-MTS, the narrower the pore size distribution and the smaller the pore volume. This decrease in pore diameter may be due to the possibility of the reaction product and/or substrates adsorbing on the support material during the modification process.

Loading of basic functional groups

The materials prepared (LP-MTS/DDA-MTS) are basic in nature and thus acid titration can be used to estimate the amount of basic groups on the surface. This is equivalent to the loading as expressed in mmol of functional groups per gram of silica. Results obtained during titration show that the loading of the amino functional group on LP-MTS material averaged at 3.3 mmol g^{-1}. This loading is higher than that on DDA-MTS (2.8 mmol g^{-1}) and that on others reported elsewhere.[29,44] The loading of amino groups onto polyamine-silica hybrids depends on the amino group source (*e.g.* monoamine, diamine) as well as the type of surfactant used.[29]

SEM analysis of the prepared materials

Scanning electron microscopy (SEM) allowed the visualization of the materials at very high magnifications. Representative electron micrographs for LP-MTS, DDA-MTS, Glu-LP-MTS,

Published on 22 August 2011 on http://pubs.rsc.org | doi:10.1039/C1CY00033K
Downloaded on 13 February 2013

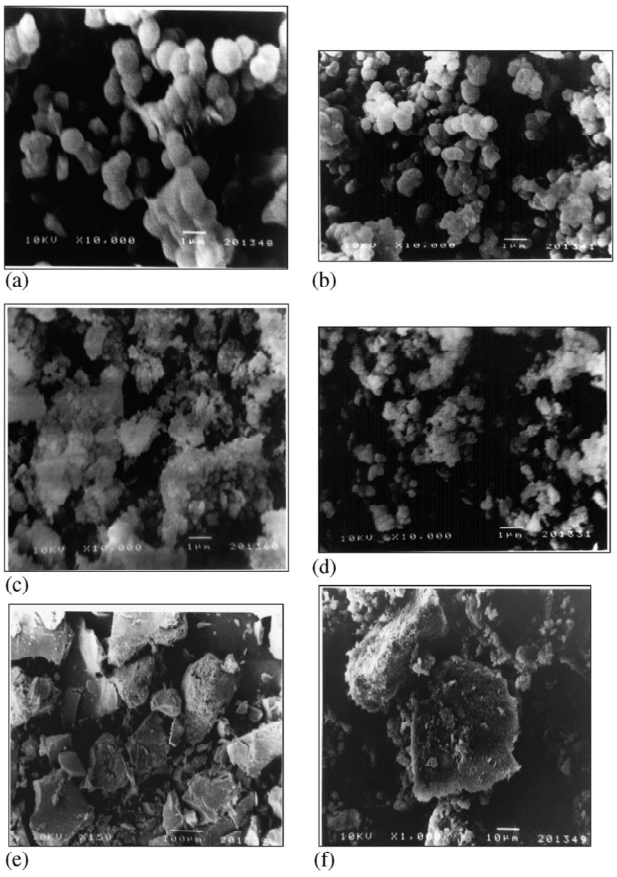


Fig. 4 SEM images of (a) LP-MTS (b) DDA-MTS (c) Glu-LP-MTS (d) Glu-DDA-MTS (e)LP-MTS covalent immobilized invertase and (f) LP-MTS cross-linked invertase.

Glu-DDA-MTS and the immobilized invertases are presented in Fig. 4(a–f).

As evident in these micrographs, there is a significant change in the morphology of the materials at different stages when LP-MTS is modified to the supported glutaraldehyde and finally to immobilized invertase enzyme. The LP-MTS (Fig. 4a) and the DDA-MTS (Fig. 4b) are comprised of roughly spherical particles that are loosely bound whereas their supported glutaraldehyde (Fig. 4c and 5d) show an aggregation of the particles. The immobilized invertase displayed a crystalline nature with rough surfaces. These rough surfaces may be a result of coagulation of enzymes after immobilization. It is likely that it is created through the breaking of supported glutaraldehyde into finer particles due to mechanical stirring and hence re-interacting to form clusters. These interactions are likely to be between the aldehyde functionalized groups of different particles or through the immobilization of enzyme molecules on two different silica particles. Moreover, the surface modifications and the effects of the mechanical stirring, creates silica particles with a rougher and finer appearance.

DRIFT analysis of LP-MTS, DDA-MTS, Glu-LP-MTS, Glu-DDA-MTS, the immobilized and free invertases

Infrared spectra were obtained using a Perkin Elmer BX FT-IR Spectrophotometer. The samples were diluted using KBr at a ratio of 9 : 1. Fig. 5 shows the spectra obtained for LP-MTS, Glu-LP-MTS, covalent immobilized invertase, free invertase and cross-linked invertase.

It is clear from the spectra that the amino group was introduced during the formation of LP-MTS due to the N–H bending mode at 1631 cm^{-1} and the C–H stretching vibration frequency observed at 2930–2940 cm^{-1} for all spectra. The corresponding simple C–H bending vibrations occur at 1408 and 1414 cm^{-1} in the spectra for the cross-linked and covalent immobilized invertases, respectively. An aldehyde group is observed on the spectrum for Glu-LP-MTS at 1744 cm^{-1} and does not appear in any other spectrum. The amine-glutaraldehyde reaction produces an imine N=C bond seen at 1634 cm^{-1} in Glu-LP-MTS. The ethylenic C=C bond formed by resonance stabilization of the imine, appears at 1551 cm^{-1} in Glu-LP-MTS spectrum and at 1561 cm^{-1} and 1566 cm^{-1} in cross-linked and covalent immobilized invertase spectra, respectively. Associated with Si–O–Si bonds are the bands, appearing in the range 1074–1081 cm^{-1} in all spectra except in the free invertase.

The spectrum of DDA-MTS (Fig. 6) shows weak C–H stretching vibration bands in the 2800–2950 cm^{-1} region and the corresponding weak H–N–H and H–C–H bending vibrations at 1640 cm^{-1} to 1550 cm^{-1}.

The spectrum of Glu-DDA-MTS shows some changes in the regions where apart from the bending vibrations due to the H–N–H and H–C–H, also a new band at about 1740 cm^{-1} attributed to a C|N stretching vibration is also observed.

Incubation time

The incubation time of the invertase enzyme and its substrate (sucrose) was varied from 10 to 70 min in order to establish the optimum incubation period. The activities obtained for free and immobilized enzymes on LP-MTS are given in Fig. 7. As seen in the Figure, as the incubation time increases, the activity increases for both free and immobilized invertases until it reaches a maximum. For both free and the immobilized invertases, the maximum activity was after 50 min incubation time.

Effect of temperature on the activities of free and immobilized invertase enzymes

The effect of temperature on the activities of free and immobilized invertase was determined by measuring the hydrolysis of sucrose at incubation temperature ranging from 25 to 60 °C. Initially, the activity of each biocatalyst increased with increasing temperature to a maximum value at which it started decreasing with increasing temperature. It is interesting to note that both the free and LP-MTS immobilized invertases exhibited a very similar temperature profile with a maximum activity between 40–43 °C (Fig. 8).

The main difference between the immobilized and the free invertases is that the immobilized invertases were stable up to slightly above 45 °C. It was noted that cross-linked invertase was slightly less stable than covalently immobilized invertase. Covalent DDA-MTS immobilized invertase generally showed very little activity compared to covalent LP-MTS immobilized and free invertase (Fig. 9).

This journal is © The Royal Society of Chemistry 2011

Published on 22 August 2011 on http://pubs.rsc.org | doi:10.1039/C1CY00033K
Downloaded on 13 February 2013

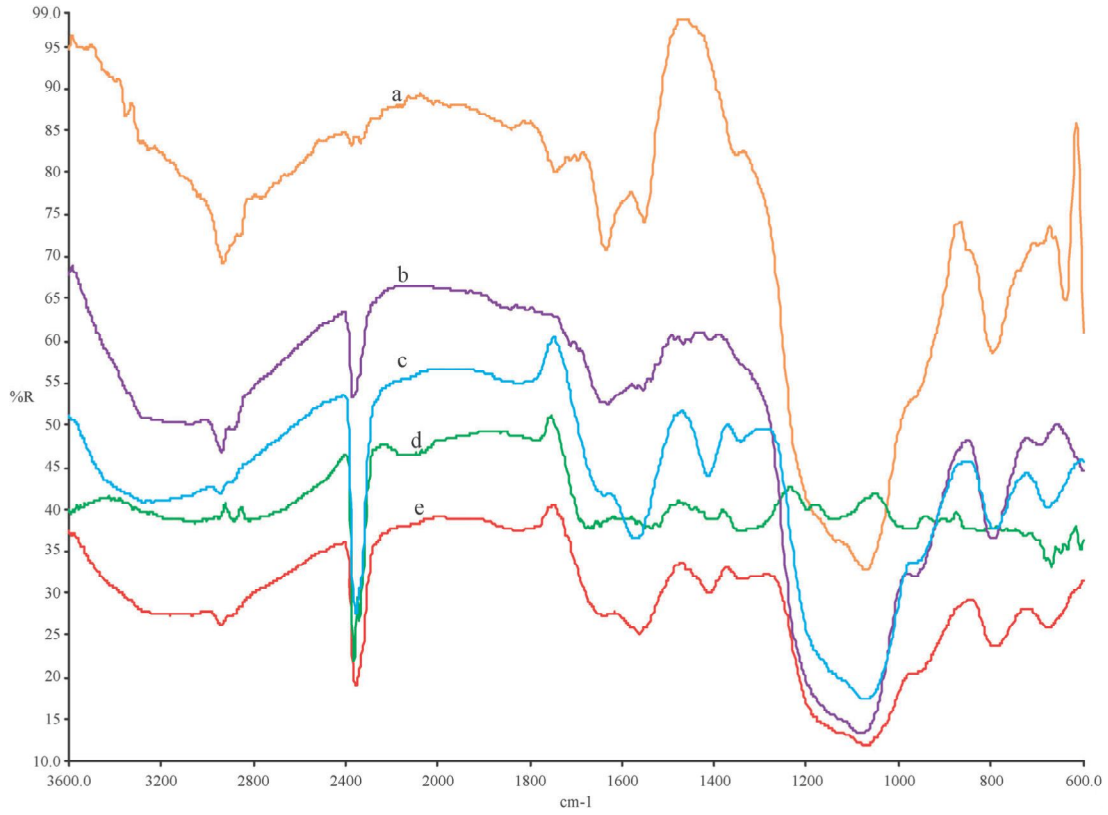


Fig. 5 FTIR spectra for (a) Glu-LP-MTS, (b) LP-MTS, (c) covalent immobilized invertase, (d) Free invertase and (e) cross-linked invertase.

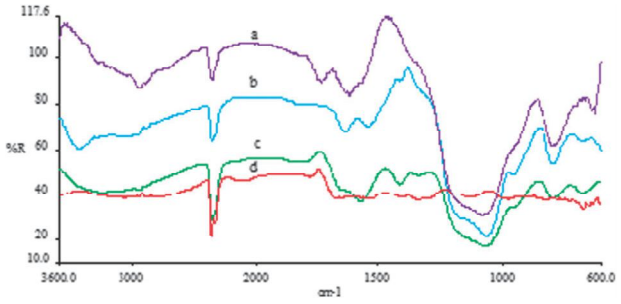


Fig. 6 FTIR spectra for (a) Glu-DDA-MTS, (b) DDA-MTS, (c) covalent immobilized invertase, and (d) Free invertase.

Reuse studies of immobilized invertases

The reuse studies of the immobilized invertase was done at 40 °C and pH 4.7 at five-day intervals over a range of 30 days so as to investigate both their shelf life and reuses (Fig. 10). As evident from the Figures, the immobilized enzymes show a high stability and they were used for up to 10 cycles in a day (Fig. 10). Both forms of the immobilized invertases were stable for about 20 days and after that, the activity started to decrease slowly (Fig. 10 cycle 7). It is evident from the reuse studies that DDA based invertase maintains its activities upon reuse whereas LP-MTS decreases in activity. This shows that in DDA supports, the enzyme is attached on the outer surface whereas in LP-MTS the enzyme is supported inside the pores and hence upon reuse pore blockage takes place reducing the access of substrate to the active sites. The blockage of catalyst pores, leading to a drop in the surface area of the catalyst from 248 m^2 g^{-1} before use to 84 m^2 g^{-1} after catalyst use, has previously been reported in non-enzymatic catalysis.[45]

Experimental

Reagents and chemicals

Yeast invertase (E.C.2.3.1.26) from baker's yeast (*S. cerevisiae*) 157 U mg^{-1}, glutaraldehyde (50%), tetraethylorthosilicate (TEOS, 98%), dodecylamine (DDA, 98%) and dinitrosalicylic acid (DNSA) were purchased from Fluka. Ethanol, potassium sodium tartrate tetrahydrate, sodium hydroxide, sodium acetate, glacial acetic acid, hydrochloric acid (assay 32%), sodium phosphate, phosphoric acid and phenol were supplied by BDH laboratory. 3-Aminopropyltrimethoxylsilane (APTMS, 97%), sucrose and glucose were supplied by Sigma-Aldrich. Cashew nut shell liquid (CNSL) was obtained from the cashew nut processing factory, Dar es Salaam, Tanzania. Distilled water was obtained from the chemistry laboratory, University of Dar es salaam. All chemicals were used as received without further purification.

Preparation of micelle templated silicas (LP-MTS or DDA-MTS)

The one pot co-condensation method was used to synthesize the LP-MTS and DDA-MTS using CNSL or DDA surfactants, respectively. The synthesis was done as described elsewhere[29] where a solution of CNSL (2.5 g) was poured into stirred aqueous ethanol (46 ml of absolute ethanol and 53 ml of distilled water) at ambient conditions of temperature and pressure.

Published on 22 August 2011 on http://pubs.rsc.org | doi:10.1039/C1CY00033K
Downloaded on 13 February 2013

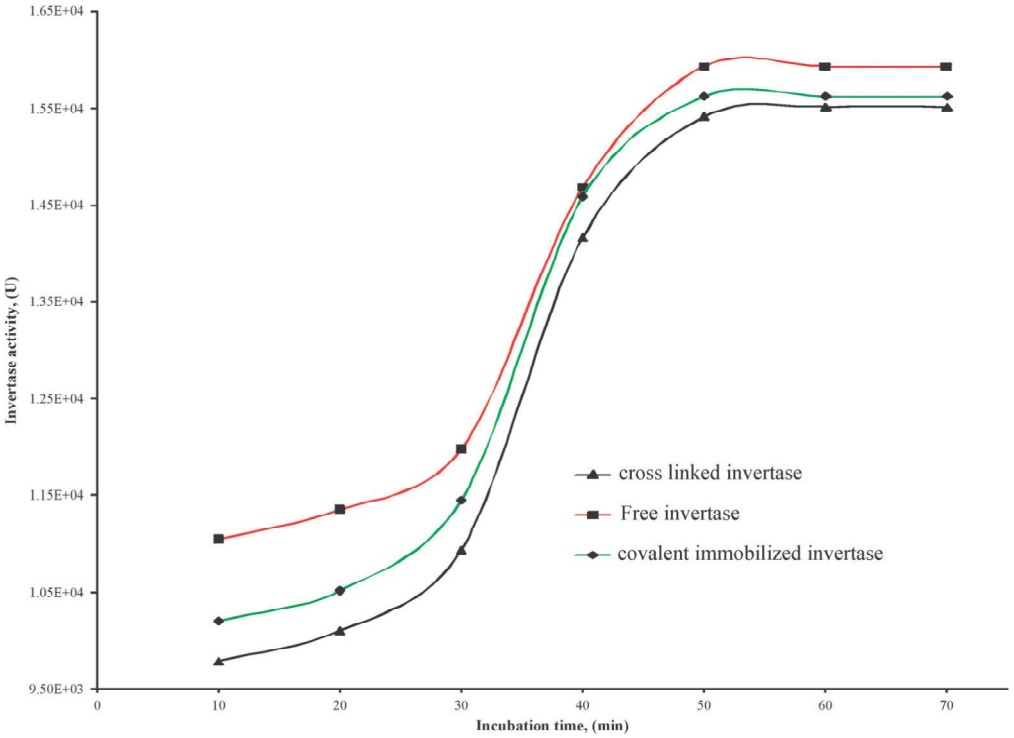


Fig. 7 Effect of incubation time on activity of invertase enzymes on LP-MTS derivatives and free invertase.

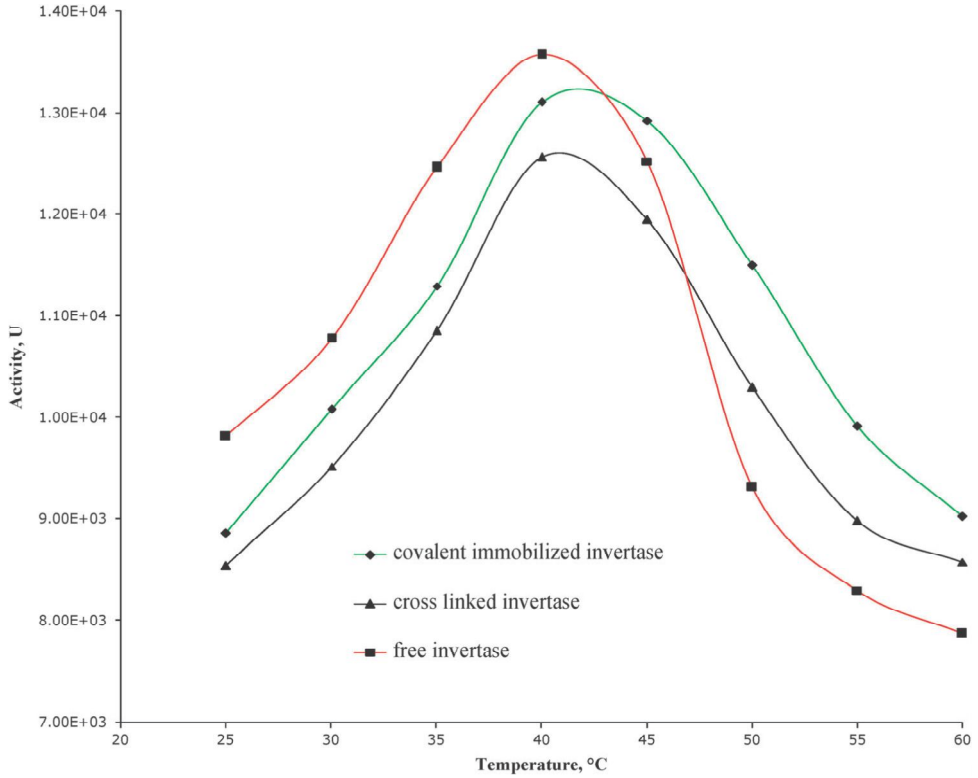


Fig. 8 Effect of temperature on the activities of free and LP-MTS immobilized invertases at pH 4.7, 50 min reaction time and 0.3 M sucrose.

To this solution, TEOS (8.3 g) and APTMS (1.9 g) were separately but simultaneously and rapidly added. The APTMS and TEOS were added at a ratio of 1 : 4 or 1 : 9. A brown solution started to thicken three minutes after addition but stirring continued for 18 h. After 18 h, the thick paste was vacuum filtered, and the brown solid was thoroughly washed with ethanol. The wet solid product was refluxed by Soxhlet using ethanol as a solvent for 10 h so as to remove the template.

This journal is © The Royal Society of Chemistry 2011

Published on 22 August 2011 on http://pubs.rsc.org | doi:10.1039/C1CY00033K
Downloaded on 13 February 2013

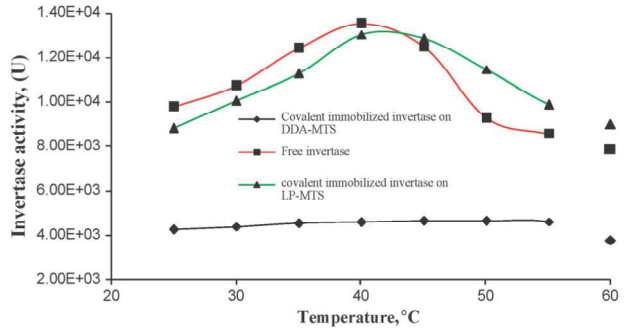


Fig. 9 Effect of temperature and type of support on the activities of free and covalent immobilized invertases.

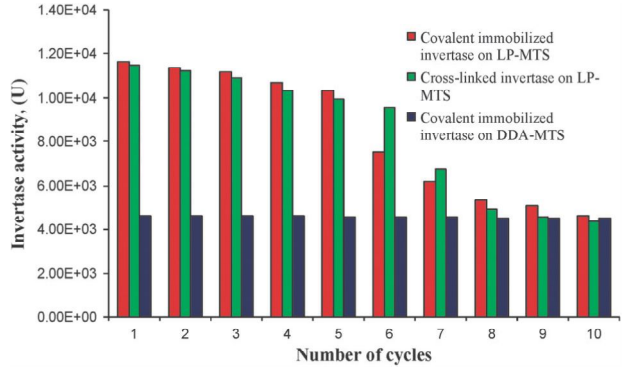
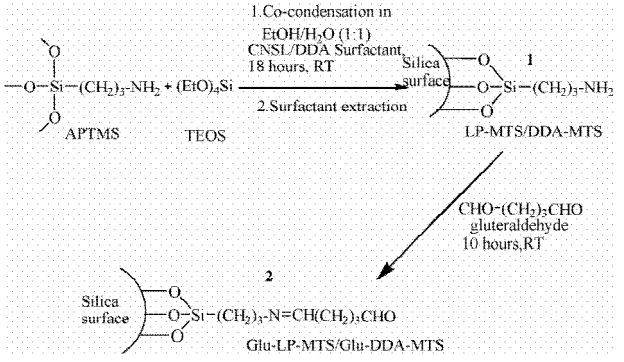


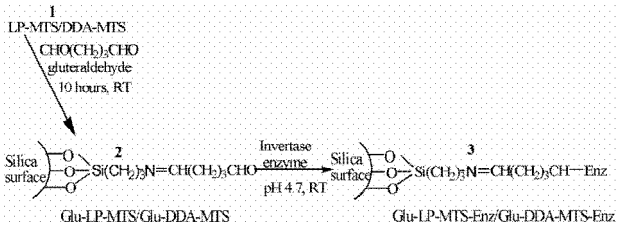
Fig. 10 Reusability of the LP-MTS and DDA-MTS immobilized invertase enzyme.

The final solid (LP-MTS) obtained after the removal of the template was dried in an oven at 100 °C for 8 h and then stored in sealed bottles and stored in a desiccator for further characterization and derivatization. The same procedure was followed for the DDA surfactant but 5 g of DDA was used and the resultant material is denoted DDA-MTS.

Functionilization of LP-MTS or DDA-MTS with glutaraldehyde. A step-wise approach (Scheme 1) was followed to give the glutaraldehyde bound support. The functionalization of the surface modified LP-MTS/DDA-MTS (**1**) was carried out by suspending 2 g of the solid LP-MTS/DDA-MTS (**1**) in 20 ml of 25% glutaraldehyde solution. The suspension was stirred at room temperature for 10 h followed by overnight incubation at 4 °C. Unbound glutaraldehyde was removed by



Scheme 1 Functionalization of LP-MTS and DDA-MTS with glutaraldehyde.



Scheme 2 Schematic representation of the covalent binding of the invertase enzyme onto Glu-LP-MTS/Glu-DDA-MTS (**2**).

vacuum filtration and then washed extensively with distilled water. The final filtered material (Glu-LP-MTS/Glu-DDA-MTS) (**2**) was dried and stored at room temperature, in a desiccator for characterization.

Immobilization of invertase on modified support material

Immobilization of invertase on the modified support material was done by using covalent and cross-linking methods.

Invertase immobilization by covalent method

In order to covalently immobilize an enzyme on LP-MTS or DDA-MTS, the amino modified silica was first reacted with glutaraldehyde and then, after extensive washing with distilled water, it was added to a buffered enzyme solution. The covalent immobilization of invertase on supported glutaraldehyde (Scheme 2) was done as described in the literature with some minor changes.[46] In a typical experiment, 0.05 g of supported glutaraldehyde (*ca.* 1 mmol) was put in 10 mL of 0.05 M sodium acetate buffer (pH 4.7), 0.03 g (*ca.* 1 mmol) of invertase enzyme was added and stirred for 5 h at room temperature followed by overnight incubation at 4 °C. The Glu-LP-MTS/Glu-DDA-MTS-invertase catalyst obtained denoted as Glu-LP-MTS-Enz/Glu-DDA-MTS-Enz (**3**) was filtered, washed and assayed for determination of immobilized invertase and characterization.

Invertase immobilization by cross-linking

Cross linking of the invertase enzyme by glutaraldehyde was done as explained elsewhere[47] with minor modifications. About 0.05 g of synthesized amine functionalized LP-MTS (**1**) in a 250 mL volumetric flask containing 10 mL of sodium acetate buffer (pH 4.7) was mixed with 0.03 g (*ca.* 1 mmol) of invertase. The mixture was then shaken in a water bath shaker for 4 h at room temperature. Thereafter, 10 mL of 25% aqueous glutaraldehyde was added and held in the water bath shaker for 3 h before keeping it at 4 °C in a refrigerator for 16 h. The immobilized invertase was then washed with excess distilled water and kept for further characterization. Unbound glutaraldehyde was removed by vacuum filtration and then washed extensively with distilled water.

Characterization of LP-MTS/DDA-MTS and supported glutaraldehyde

The LP-MTS/DDA-MTS support and supported glutaraldehyde (Glu-LP-MTS/Glu-DDA-MTS) were characterized by standard techniques including nitrogen physisorption, DRIFT, acid titration and SEM.

Published on 22 August 2011 on http://pubs.rsc.org | doi:10.1039/C1CY00033K
Downloaded on 13 February 2013

Activity studies of free and immobilized invertases

The activity of free and immobilized invertase was done by measuring the rate of formation of reduced sugars during the hydrolysis of sucrose (0.3 M) in 0.05 M sodium acetate buffer (pH 4.7) at 40 °C. The optimum incubation time for the enzyme and its substrate was found to be 50 min and 10 min for the LP-MTS and DDA-MTS derivatives respectively. All other experiments were done under these conditions except when a specific parameter was under investigation. The total reduced sugar produced was measured calorimetrically in triplicate, using a dinitrosalicylic acid (DNSA) method. The activities of the free and immobilized invertases were performed at different temperatures, pH values and substrate concentration to study their effects on the enzymes. A unit of invertase activity (U) corresponds to the amount of enzyme hydrolyzing sucrose at 1 μmol min^{-1}.

The activities of the free and immobilized invertases were determined according to the method described in the literature[48] with some minor modifications. The free invertase (*ca.* 1 mmol) and immobilized invertase (*ca.* 1 mmol) were each incubated with a sucrose solution (0.3 M) in 10 mL of 0.05 M sodium acetate buffer (pH 4.7) at a temperature range of 25 °C to 60 °C for 50 min in order to investigate the effect of temperature and was followed colorimetrically at A_{430} with a light path length of 1 cm. This was done in triplicate and the reducing sugar produced was determined by the DNSA method described elsewhere.[49] The same procedure was followed but at the pH range 1–9 at 40 °C to investigate the effect of pH. Different concentrations of sucrose solution (0.1–0.6 M) in 10 mL of 0.05 M sodium acetate buffer were also done so as to establish the effect of substrate concentration on invertase activity.

The reusability of immobilized invertases

The reusability of the immobilized invertases was done as described in the literature.[50] The activity of a given immobilized invertase was tested several times in a day and after each test, the immobilized invertase was washed with sodium acetate buffer (pH 4.7) before being re-tested. The insoluble invertase was stored in a refrigerator at 4 °C in 10 mL of sodium acetate (0.05 M) buffer (pH 4.7). The storage tests were done in the interval of five days and were done for a period of one month. There was a notable decrease in the activity after one month of shelf life. The thermal and pH stability of immobilized invertase was tested using a similar procedure but at different temperatures (from 25 °C to 60 °C) and pH values ranging from 1 to 9.

Conclusion

A new mesoporous micelle-templated silica route for enzyme immobilization has been developed using large pore micelle templated silica prepared using a natural and renewable surfactant, a CNSL template. The results of this study show that large pore micelle templated silicas (LP-MTS) derived from a CNSL templates have large pore diameters up to 25 nm depending the condition of preparation. These materials are suitable for immobilization of biomolecules compared to dodecylamine based micelle templated silica, DDA-MTS. In general large pore micelle templated silicas prepared using CNSL templates produced materials with larger pore dimensions than those of DDA derivatives. The LP-MTS immobilized enzymes are active, stable and can be stored for over twenty days and reused without significant loss of activity.

Acknowledgements

Sida-Sarec and Faculty of Science-University of Dar es Salaam under the Capacity Building in Material Science and Solar Energy Project is acknowledged for sponsoring this work.

References

1 J. M. Woodley, *Trends Biotechnol.*, 2008, **26**, 321–327.
2 L. Que and W. B. Tolman, *Nature*, 2008, **455**, 333–340.
3 J. S. Dordick and A. Freeman, *Curr. Opin. Biotechnol.*, 2006, **17**, 559–561.
4 H. E. Schoemaker, D. Mink and M. G. Wubbolts, *Science*, 2003, **299**, 1694–1697.
5 C.-H. Lee, T.-S. Lin and C.-Y. Mou, *NanoToday*, 2009, **4**, 165–179.
6 S. Pankel and M. Wubbolts, *Curr. Opin. Chem. Biol.*, 2005, **9**, 188–194.
7 Y. Li, G. Zhou, W. Qiao and Y. Wang, *Mater. Sci. Eng., B*, 2009, **162**, 120–126.
8 H. H. P. Yiu, P. A. Wright and N. P. Botting, *J. Mol. Catal. B: Enzym.*, 2001, **15**, 81.
9 H. H. P. Yiu and P. A. Wright, *J. Mater. Chem.*, 2005, **15**, 3690–3700.
10 M. Mureseanu, A. Galarneau, G. Renard and F. A. Fajula, *Langmuir*, 2005, **21**, 4648–4655.
11 A. L. Crumbliss, J. Stonehuerner, R. W. Henkens, J. P. O'Daly and J. Zhao, *New J. Chem.*, 1994, **18**, 327–339.
12 W. Tischer and V. Kasche, *Trends Biotechnol.*, 1999, **17**, 326–334.
13 T. Nagayasu, M. Miyanaga, T. Tanaka, T. Sakiyama and K. Nakanishi, *Biotechnol. Bioeng.*, 1994, **43**, 1108–1117.
14 Y.-F. Chang and D.-F. Tai, *Tetrahedron: Asymmetry*, 2001, **12**, 177–179.
15 J. Livage, T. Coradin and C. Roux, *J. Phys.: Condens. Matter*, 2001, **13**, 673–691.
16 C. Lei, T. A Soares, Y. Shin, J. Liu and E. J Ackerman, *Nanotechnology*, 2008, **19**, 125102–125111.
17 H. Ikemoto, Q. Chi and J. Ulstrup, *J. Phys. Chem. C*, 2010, **114**, 16174–16180.
18 X. Bai, Z. Ye, Y. Li, L. Yang, Y. Qu and X. Yang, *Biochem. Eng. J.*, 2010, **49**, 264.
19 X. S. Zhao, Xiao Y. Bao, W. Guo and F. Y. Lee, *Mater. Today*, 2006, **9**, 32–39.
20 K.-C. Kao, C.-H. Lee, T.-S. Lin and C.-Y. Mou, *J. Mater. Chem.*, 2010, **20**, 4653–4662.
21 M. Vallet-Regi, F. Balas and D. Arcos, *Angew. Chem., Int. Ed.*, 2007, **46**, 7548.
22 I. I. Slowing, J. L. Vivero-Escoto, C. W. Wu and V. S. Lin, *Adv. Drug Delivery Rev.*, 2008, **80**, 1278–1288.
23 J. M. Rosenholm, C. Sahlgren and M. Lindèn, *Nanoscale*, 2010, **2**, 1870–1883.
24 A. Macario, M. Moliner, A. Corma and G. Giordano, *Microporous Mesoporous Mater.*, 2009, **118**, 334–340.
25 A. Corma, V. Fornes and F. Rey, *Adv. Mater.*, 2002, **14**, 71–74.
26 E. Serra, E. Díez, I. Díaz and R. M. Blanco, *Microporous Mesoporous Mater.*, 2010, **132**, 487–493.
27 E. Serra, A. Mayoral, Y. Sakamoto, R. M. Blanco and I. Díaz, *Microporous Mesoporous Mater.*, 2008, **114**, 201–213.
28 S. Urrego, E. Serra, V. Alfredsson, R. M. Blanco and I. Díaz, *Microporous Mesoporous Mater.*, 2010, **129**, 173–178.
29 A. Hilonga, J. E. G. Mdoe and L. L. Mkayula, *Int. J. BioChemi-Physics*, 2009, **17**, 26–31.
30 P. S. J. Cheetham, *The applications of enzymes in industry. Handbook of Enzyme Biotechnology*, Ellis, London, 1995, p. 420.

This journal is © The Royal Society of Chemistry 2011

View Article Online

31 K. Nakane, T. Ogihara, N. Ogata and Y. Kurokawa, *J. Appl. Polym. Sci.*, 2001, **81**, 2084.
32 G. Bayramoglu, S. Akgol, A. Bulut, A. Denizli and M. Y. Arica, *Biochem. Eng. J.*, 2003, **14**(2), 117.
33 E. H. Mansour and F. M. Dawoud, *J. Sci. Food Agric.*, 2003, **83**(5), 446.
34 S. Kiralp, L. Toppare and Y. Tagci, *Synth. Met.*, 2003, **135**(1–3), 79.
35 C. Airoldi and O. A. C. Monteiro, *Polym. Bull.*, 2003, **50**(1–2), 61.
36 M. Fuentes, J. V. Maquiese, B. C. C. Pessela, O. Abian, R. Fernandez-Lafuente, C. Mateo and J. M. Guisan, *Biotechnol. Prog.*, 2004, **20**(1), 284.
37 G. Sanjay and S. Sugunan, *Catal. Commun.*, 2005, **6**, 81.
38 S. A. Camperi and M. Grasselli, *Process Biochem.*, 2004, **39**(8), 1017.
39 I. S. Khatib and R. V. Parish, *J. Organomet. Chem.*, 1989, **369**, 9–16.
40 J. E. G. Mdoe and D. J. Macquarrie, *Int. J. Sci. Res.*, 2005, **14**, 19–25.
41 E. F. Vansant, P. Van Der Voort and K. C. Vranken, in *Characterisation and Chemical Modification of the Silica Surface*, Elsevier, Amsterdam, 1995.
42 S. J. Gregg and K. S. W. Sing, *Adsorption, Surface Area and Porosity*, Academic Press, London, 1982.
43 S. Brunauer, P. H. Emmett and E. Teller, *J. Am. Chem. Soc.*, 1938, **60**, 309.
44 E. B. Mubofu, *PhD Thesis*, University of York, UK, 2001.
45 J. H. Clark, D. J. Macquarrie and E. B. Mubofu, *Green Chem.*, 2000, **2**, 53–56.
46 A. E. David, N. S. Wang, V. C. Yang and A. J. Yang, *J. Biotechnol.*, 2006, **125**, 395–407.
47 W. L. Stanley and A. C. Oslon, *J. Food Sci.*, 1974, **39**, 660–666.
48 *Immobilized cells and enzymes, A practical approach*, ed. J. Woodward, IRL Press, Oxford, Washingtone DC, 1985, p. 3; Bergmeyer, *Methods of Enzymatic Analysis*, 1974, **I**, 450–451.
49 G. L. Miller, *Anal. Chem.*, 1959, **31**, 426.
50 A. Nighojkar, S. Srivasttva and A. Kumar, *Indian J. Exp. Biol.*, 1996, **34**, 1248–1253.

Published on 22 August 2011 on http://pubs.rsc.org | doi:10.1039/C1CY00033K
Downloaded on 13 February 2013